

Selective Antiproliferative Activity of an Oleandrigenin-Derived Androstane in Prostate Cancer Cells

Martina Metz^{a,b}, Jana Steigerová^{a,b*}, Marie Kvasnicová^{c,d}, Zlata Rychnovská^a, Tereza Kašparová^a, Tereza Štenclová^c, Lucie Rárová^d, Karol Michalak^e, Jerzy Wicha^e, Miroslav Strnad^d

^aDepartment of Clinical and Molecular Pathology, Faculty of Medicine and Dentistry, Palacký University, Hněvotínská 3, 775 15 Olomouc, Czech Republic

^bInstitute of Molecular and Translation Medicine, Faculty of Medicine and Dentistry, Palacký University and Faculty Hospital in Olomouc, Puškinova 6, 775 20 Olomouc, Czech Republic

***Corresponding author.**

Jana Steigerova, Department of Clinical and Molecular Pathology, Faculty of Medicine and Dentistry, Palacký University and University Hospital, Hněvotínská 3, 77515 Olomouc Czech Republic, Tel.: +420 585 639 541; email: jana.steigerova@upol.cz

Supplementary Information

Tab. S1 Chemical names of tested cardiac glycoside derivatives

Compound No.	Chemical name
1	Oleandrogenin
2	3-O-Acetyl oleandrogenin
3	16 β -Acetoxy-17 β -(Furyl-3)-5 β , 14 β -androstan-3 β , 14-diol
4	17 β -(Furyl-3)-5 β , 14 β -androstan-3 β , 14, 16 β -triol
5	17 β -(Furyl-3)-3 β -tertbutyldimethylsilyloxy-5 β , 14 β -androstan-14, 16 β -diol
6	3-O-Methyl-5 α -oleandrogenin
7	16 β -Acetoxy-17 β -(5H-furan-2-on-3-yl)-3 β -methoxy-5 α -androst-14-ene
8	17 β -(5H-Furan-2-on-3-yl)-3 β -methoxy-5 α -androstan-14, 16-diene
9	16 β -Acetoxy-14, 15 β -epoxy-3 β -methoxy-5 α -card-22-enolide
10	16 β -Acetoxy-14, 15 α -epoxy-17 β -(5H-furan-2-on-3-yl)-3 β -methoxy-5 α , 14 α -androstan

Tab. S2 List of primary (A) and secondary (B) antibodies used in experiments.

A. Primary antibodies

Target	Clone	Type	Host	Dilution	Manufacturer
AR	N-20	polyclonal	rabbit	1:200/1:50*	Santa Cruz Biotechnology
pAR (Ser81)	–	monoclonal	rabbit	1:500	Sigma-Aldrich
Bcl-2	100	monoclonal	mouse	1:500	Biogenex
Bcl-X _L	H-62	monoclonal	rabbit	1:100	Novocastra
Bid	B-3	monoclonal	mouse	1:100	Santa Cruz Biotechnology
β -actin	C4	monoclonal	mouse	1:1000	Santa Cruz Biotechnology
Caspase-3	Asp175	polyclonal	rabbit	1:1000	Cell Signaling Technology
CDK4	DCS-31	monoclonal	mouse	1:200	Santa Cruz Biotechnology
Cyclin A	H432	polyclonal	rabbit	1:200	Santa Cruz Biotechnology
Cyclin B1	GNS1	monoclonal	mouse	1:1000	Cell Signaling Technology
Cyclin D ₁	72-13G	monoclonal	rabbit	1:100	Santa Cruz Biotechnology
Cyclin E	HE12	monoclonal	mouse	1:200	Santa Cruz Biotechnology
ER- α	D-12	monoclonal	mouse	1:200/1:50*	Novocastra
ER- β	H-150	monoclonal	rabbit	1:100/1:50*	Santa Cruz Biotechnology
MCM-7	141.7	monoclonal	mouse	1:500	Sigma-Aldrich
Nkx3.1	D2Y1A	monoclonal	rabbit	1:500	Cell Signaling Technology
p16	JC8	monoclonal	mouse	1:100	Santa Cruz Biotechnology
p21	12D1	monoclonal	rabbit	1:1000	Cell Signaling Technology
p53	DO1	monoclonal	mouse	1:500	Invitrogen
PSA/KLK3	D6B1	monoclonal	rabbit	1:250	Cell Signaling Technology
Rb	4H1	monoclonal	mouse	1:1000	Cell Signaling Technology
pRb (S807/811)	S807/811	polyclonal	rabbit	1:1000	Cell Signaling Technology
Src	-	monoclonal	mouse	1:1000	Cell Signaling Technology

* dilution for immunofluorescence

B. Secondary antibodies

Target	Description	Dilution	Manufacturer
HRP-conjugated goat anti-mouse IgG	Goat anti-mouse IgG (H+L), HRP conjugated	1:6000	Cell Signaling Technology
HRP-conjugated goat anti-rabbit IgG	Goat anti-rabbit IgG (H+L), HRP conjugated	1:6000	Cell Signaling Technology
Alexa Fluor 594-conjugated goat anti-mouse IgG	Goat anti-mouse IgG (H+L), Alexa Fluor 594 conjugated	1:1000	Thermo Fisher Scientific (Life Technologies)

Target	Description	Dilution	Manufacturer
Alexa Fluor 488-conjugated goat anti-rabbit IgG	Goat anti-rabbit IgG (H+L), Alexa Fluor 488 conjugated	1:1000	Thermo Fisher Scientific (Life Technologies)

Fig. S1 Structures of tested cardiac glycoside derivatives.

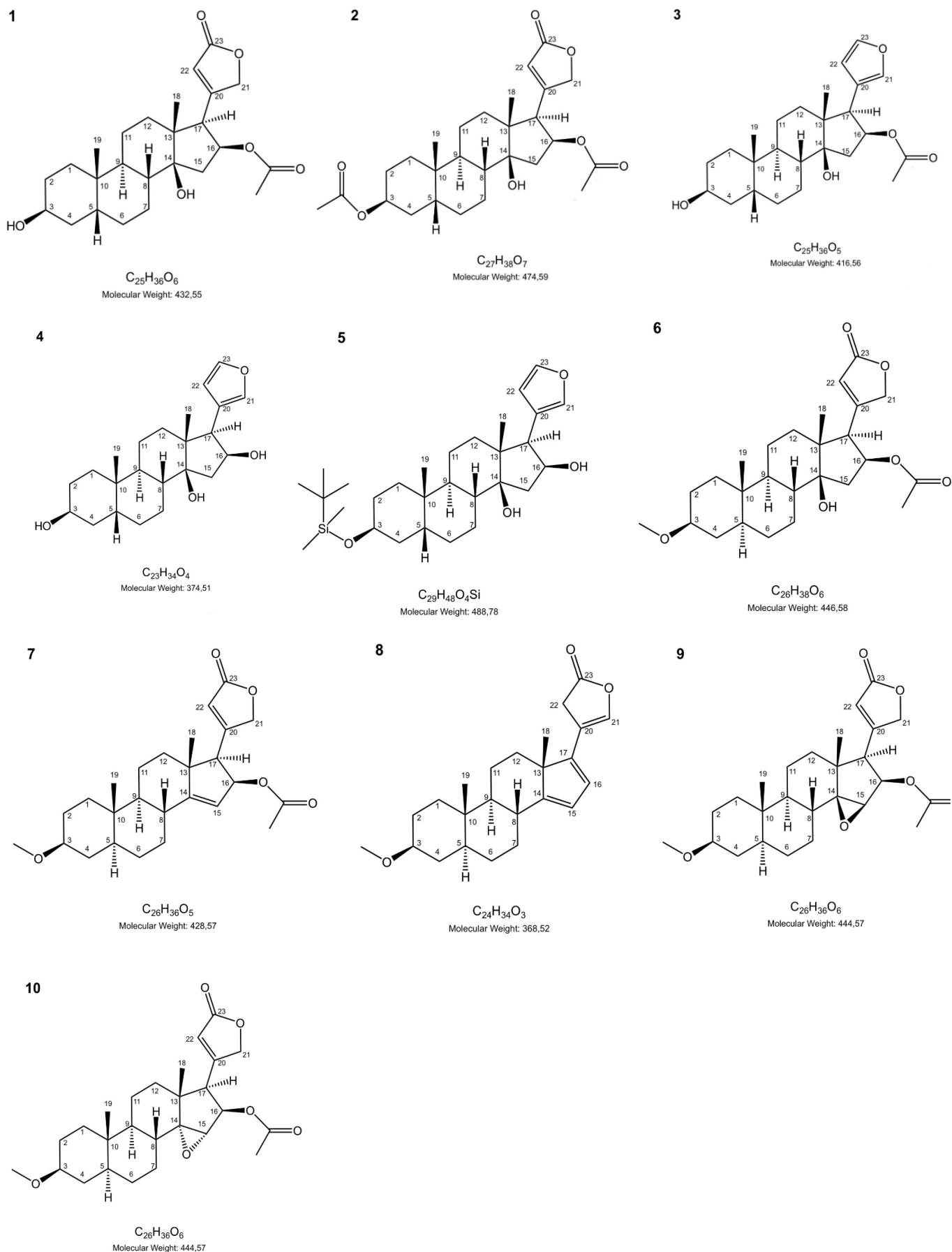


Fig. S2 Representative bright-field images of untreated control and compound **10**-treated RWPE-1, DU-145 and LNCaP cells after 24 and 48 h of treatment at the IC₅₀ concentration. Magnification, 200×.

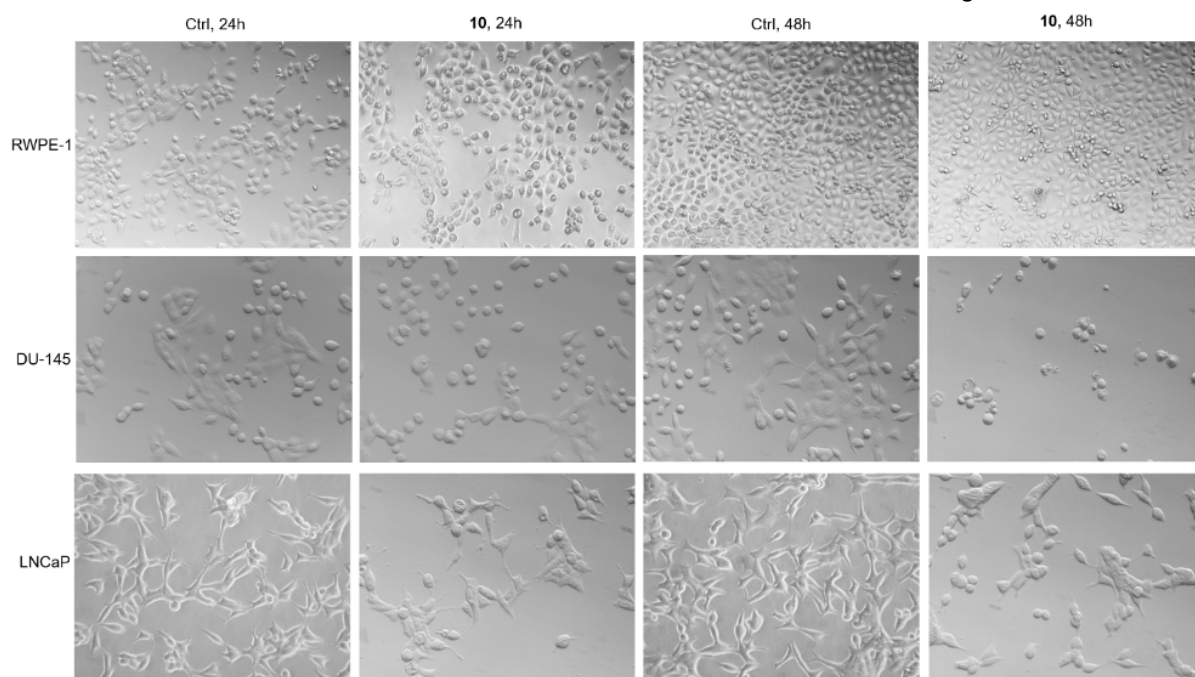
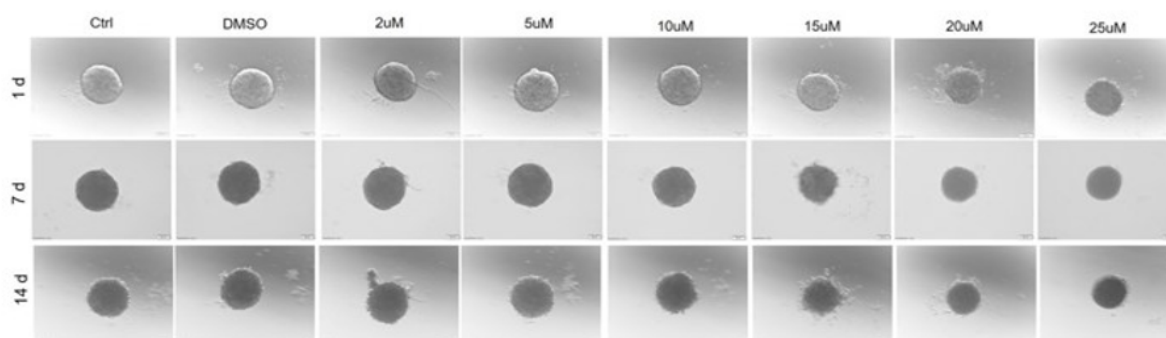


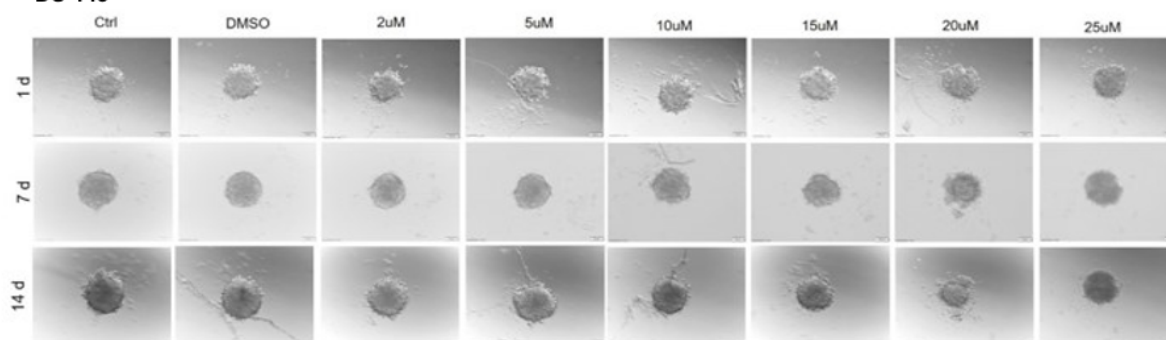
Fig. S3 Time-dependent effect (1–14 days) of compound **10** (2–25 μM) on the cellular integrity of RWPE-1, DU-145 and LNCaP spheroids. **(A)** Representative bright-field images of untreated control and compound **10**-treated spheroids after 1, 7 and 14 days of treatment at all tested concentrations (2–25 μM). Magnification, 100 $\times$. Quantification of spheroid area in **(B)** RWPE-1, **(C)** DU-145 and **(D)** LNCaP cells. Data are expressed relative to untreated controls. Asterisks indicate significant differences compared with the corresponding control (*) $p < 0.05$, (**) $p < 0.01$.

A

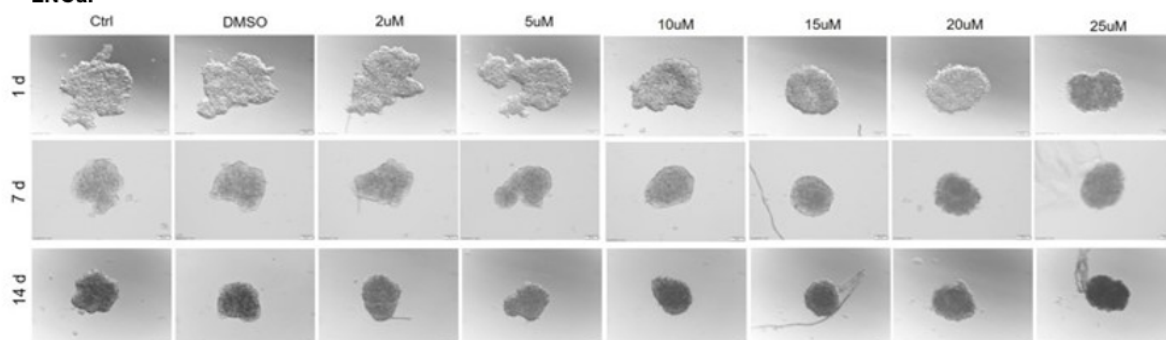
RWPE-1



DU-145



LNCaP



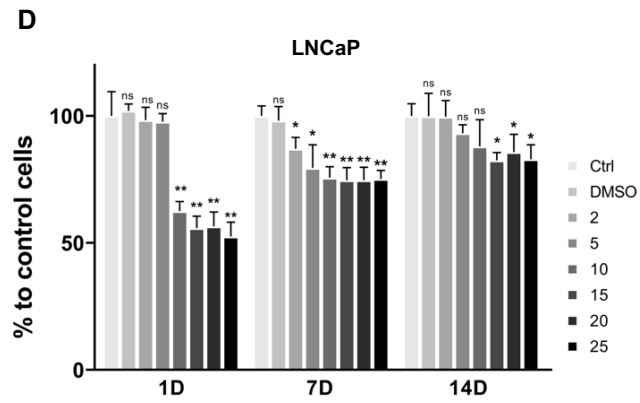
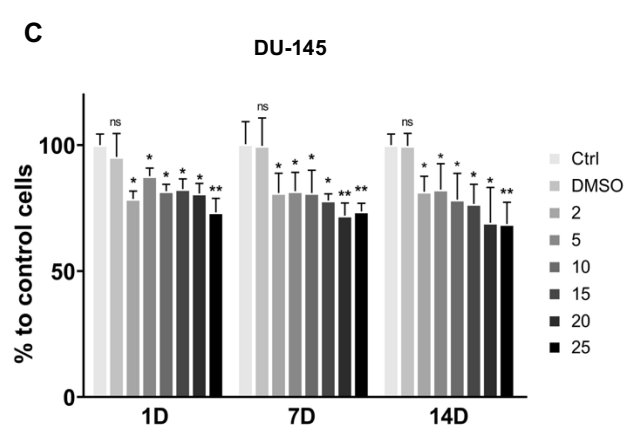


Fig. S4 Complete immunofluorescence images of nuclear steroid receptors in DU-145 and LNCaP cells treated with compound **10** at the IC₅₀ concentration for 24 h. ER- α and ER- β are shown for DU-145 cells, whereas AR, ER- α and ER- β are shown for LNCaP cells. Individual fluorescence channels (green), DAPI (blue), and the corresponding merged images are shown for each treatment condition. Magnification, 400 $\times$.

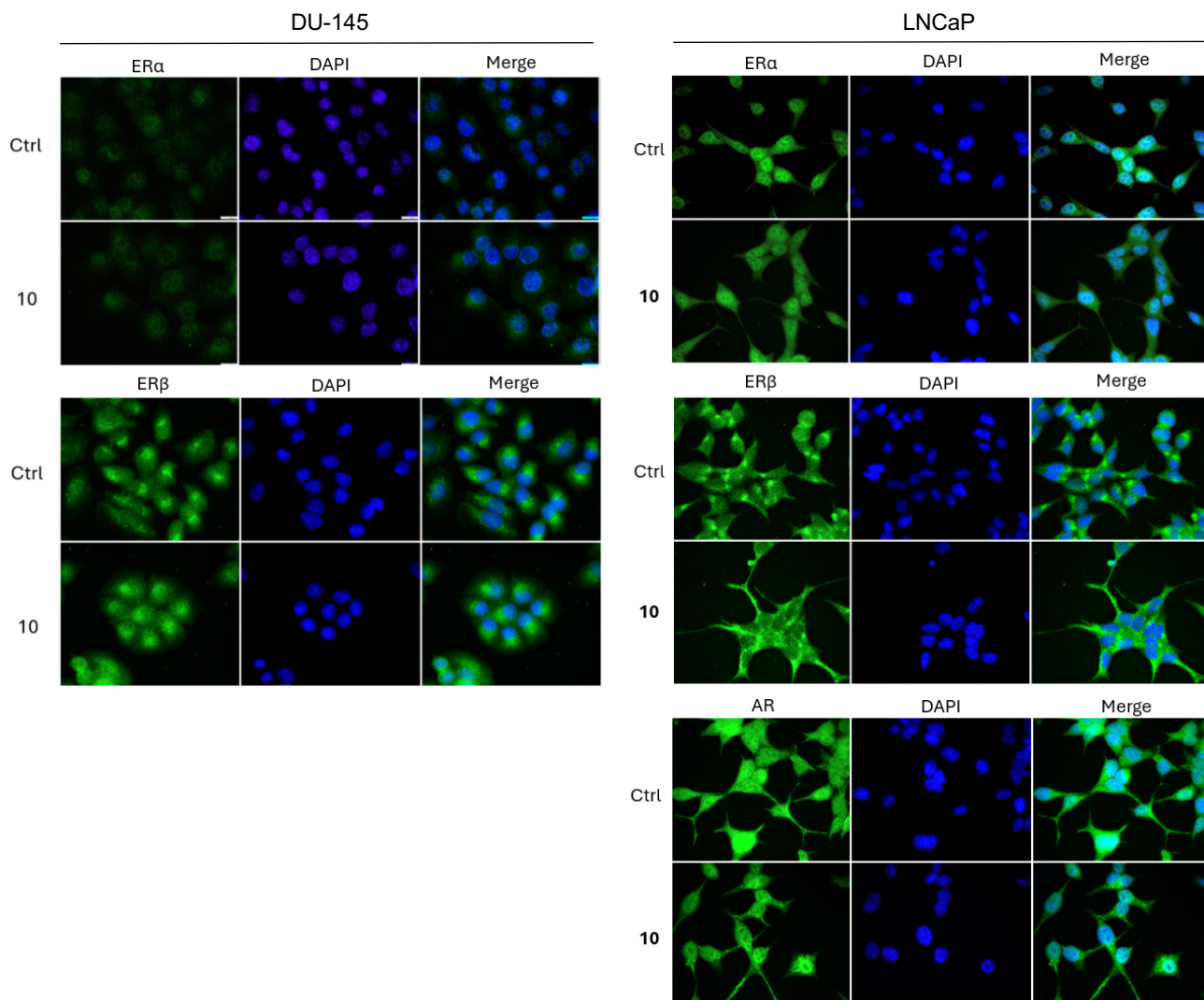
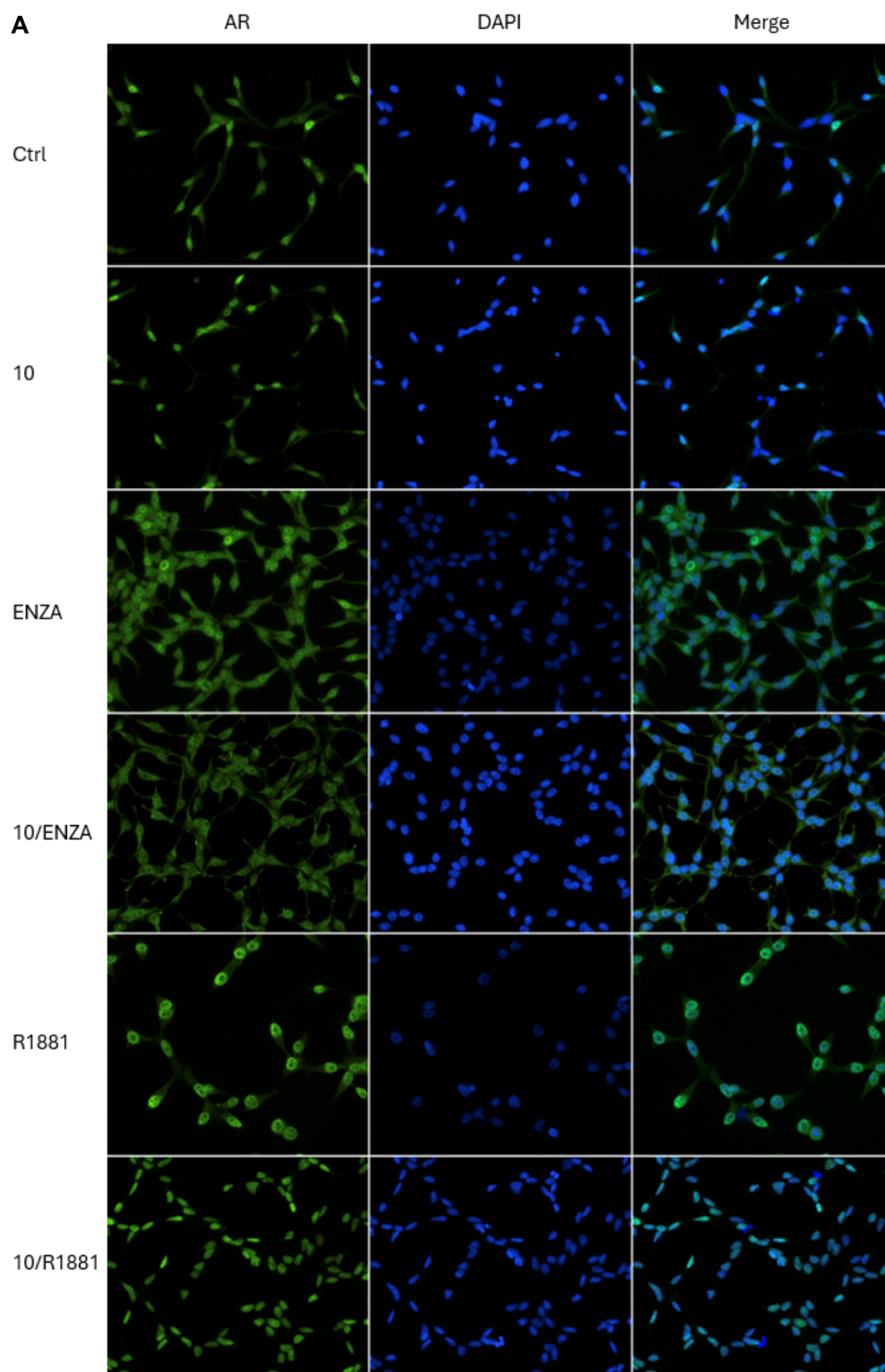


Fig. S5 Complete immunofluorescence images of AR and phosphorylated AR (Ser81) in LNCaP cells treated with compound **10** at the IC₅₀ concentration, R1881 (1 nM), enzalutamide (1 μ M), or their combinations for 24 h in charcoal-stripped serum medium. Individual fluorescence channels for AR or pAR (Ser81) (green), DAPI (blue), and the corresponding merged images are shown for each treatment condition. Magnification, 200 $\times$.



B