

Supplementary Results

A novel long non-coding RNA regulates the integrin, ITGA2 in breast cancer

Breast Cancer Research and Treatment

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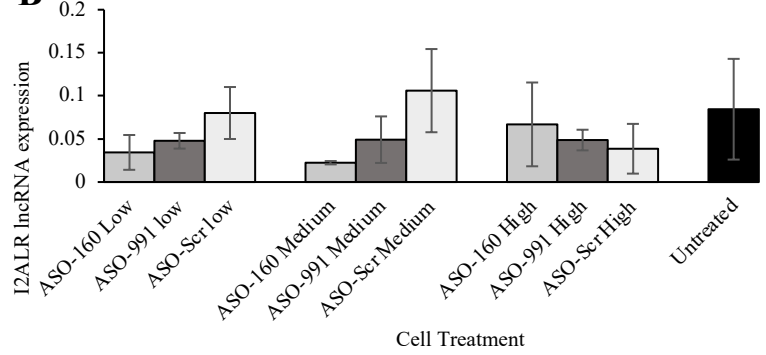
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Supplementary Results Figure 1

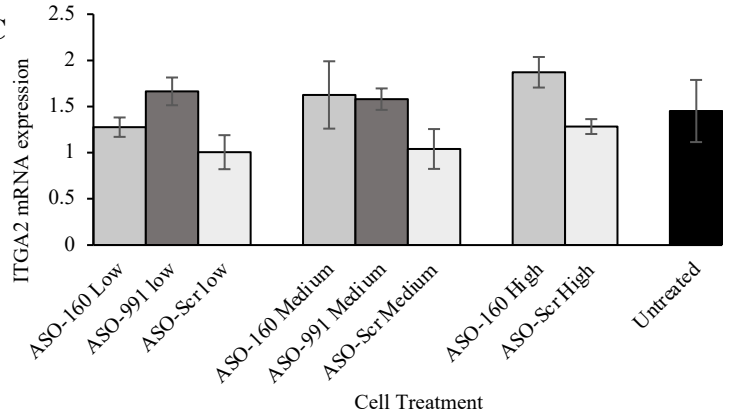
A

Exon 1 >hg38 range=chr5:52990090-52990278
ATTGCGGCATCTAGGATCCAGAAGTGAAGAAGTCTTCTAGCCTAAGT
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GAAGGAAGAATTCCCTCTGCCAAGTATTAAAGTGGCGCTTGGTTTATTC
CAAGTCCCCCGGGAAGAACACAGCGCTTCGTCCTCAA
Exon 2 >hg38 range=chr5:52988204-52988276
ATCTTCATGGAATGCTACTATGAGCTGAATGGGATGCCTGAGATACTGCC
TCTAGGAAGCAAAGTGGCTCAAG
Exon 3 >hg38 range=chr5:52978252-52978399
AGGCTGGCTGGCAAGATGGCCGAATAGGAACAGCTCCAGTCTGCAGCTCC
TAGTGAGATCAACGCCAAAGCGGGTGACTTCTGCATTTCCAACTGAGAT
ACCCAGCTCATCTCACTGGGACTGGTTAGACA GTGGGTGCAGCCCATG
Exon 4 >hg38 range=chr5:52948825-52948907
AGGTACAAGGCCTGAAAGAACCTATCCAATATCAGCTCTGGTTTCTCTC
TTCTCCTGCCAGCGGTGTTCTAAAACCATAGAG
Exon 5 >hg38 range=chr5:52932419-52932536
ATTGAGAAGACTGATCAGCTTCCACAGGATGAGAACGATGACCAAGAAAT
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CTTCAACTTGCAATTCGGG
U3R >hg38 range=chr5:52931419-52932418
aattaagactggacagcaaatgcaatccgggtatcactaacacctaact
ttgggttatttagtataatggttctcaaacatttaggatgcatgagaatta
cctggaaggcttggttaaccccatcctccgagtttctgatttgtaggtt
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acttoatattttaccgggatagaaatgtttactacattatcagaagggt
ttcaggaggtttctgctgtcactagaaaa **acacgggcacattcagacaa**
accattaaaatcttgactatttaaagacatttctcaagtgtttgaaagaga
agcttatcctgggaacta **ataaaa**gcgatgtaagagtataatttttaat
■-poly(A)

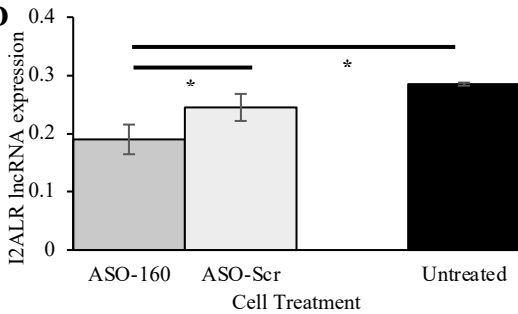
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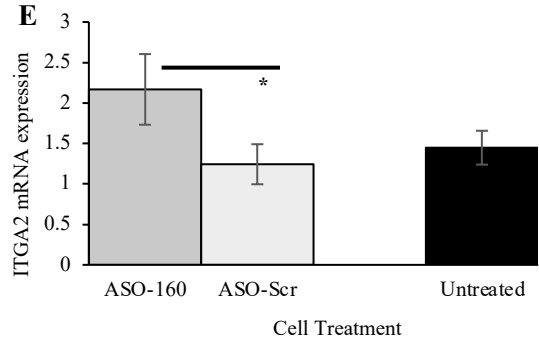
C



D



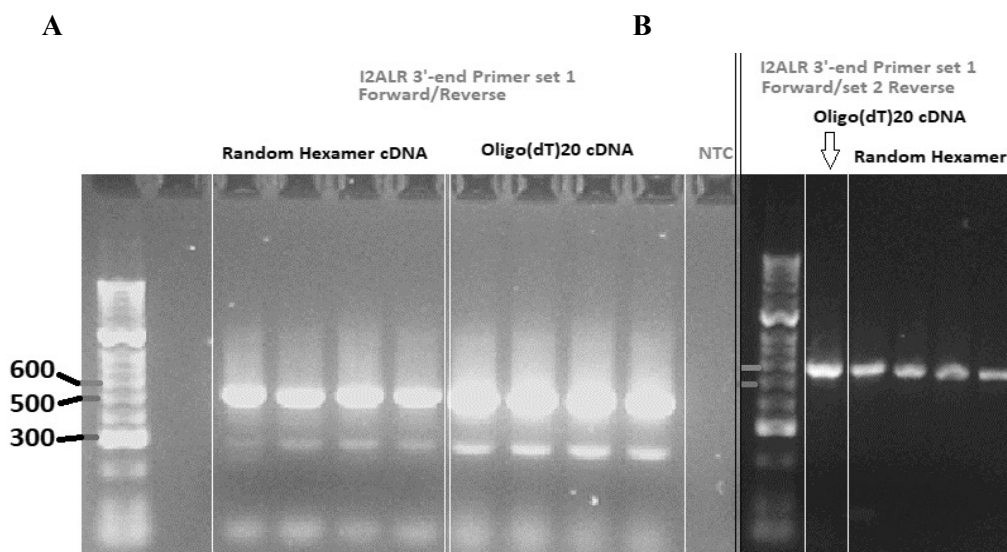
E



Supplementary Fig. 1. (A) Sequence for *I2ALR* combined exons (*ENST00000505701.5* + *ENST00000503559.1*, orange) and downstream U3R including poly-A signal (blue, signal highlighted red), the recognition sites for ASOs (ASO-160 and ASO-991) are highlighted light-blue and yellow respectively, with primers used to amplify the U3R underlined. (B) MDA-MB-453 cell *I2ALR* expression following ASO-160 or ASO-991 transfection after 24-hours at 10 nM (low), 25 nM (medium) and 50 nM (high). (C) *ITGA2* expression after 24-hours transfection at ASO concentrations low, medium, and high. (D) *I2ALR* expression in MD-MB-453 cells at 48-hours following transfection at 10nM ASO-160 (E) *ITGA2* expression after 48-hour following transfection with ASO-160 at 10nM. (B, C) n = 1 biological replicates, statistical significance not calculated; (D, E) n = 3 biological replicates, expression absolute copy number normalized to *GAPDH*, $\pm$ SEM. Statistical significance (* $P \leq 0.05$, ** $P \leq 0.01$, *** $P \leq 0.001$).

Supplementary Results Figure 2 : Characterisation of I2LAR

Sequence analysis of the most 3' *I2ALR* exon revealed two putative poly-adenylation motifs. A classical motif (*vide* Proudfoot, 2011) with 5'-AATAAA signal (469–474 bp downstream from the exon) with a 5'-CA cleavage site 26 bp further downstream. In addition, a poly-A enhancing G/U rich region was identified 3' to the cleavage site and a uracil rich region 5' to the motif (Figure 5 B). A non-classical poly-A signal was identified 404–429 bp downstream from the exon. To characterise the 3'- poly-A signal, a DNA fragment was amplified employing a primer anchored in the last known exon to several sites adjacent to the poly-A signals. A 511 bp fragment was amplified ('*I2ALR* 3'-end primer set 1') from cDNA generated using oligo(dT)₂₀ (Supplementary Results Figure 2A). Sanger sequencing indicated that exon 5 of *I2ALR* extended at least 405 bp further downstream without additional splicing (Figure 5 B). To distinguish between the classical and non-classical poly-A motifs, the '*I2ALR* 3'-end set 1' forward primer was combined with the '*I2ALR* 3'-end set 2' reverse primer (primer sequences Supplementary Methods Table 2) so that the reverse primer binding site was located between the two motifs (as shown Figure 5 B). This was performed using cDNA generated with oligo(dT)₂₀ or random hexamers, and a product was generated, matching the expected 565 bp size (Supplementary Results Figure 2 B, below), suggesting that the classical poly-A signal was functional, and *I2ALR* was polyadenylated with its 3' most exon 619 bp in length (designated as U3R). The *I2ALR* transcription start site (TSS) and promoter are located 500–600 bp downstream from the *ITGA2* first exon, however boundaries of the promoter elements and precise lncRNA TSS are not defined.



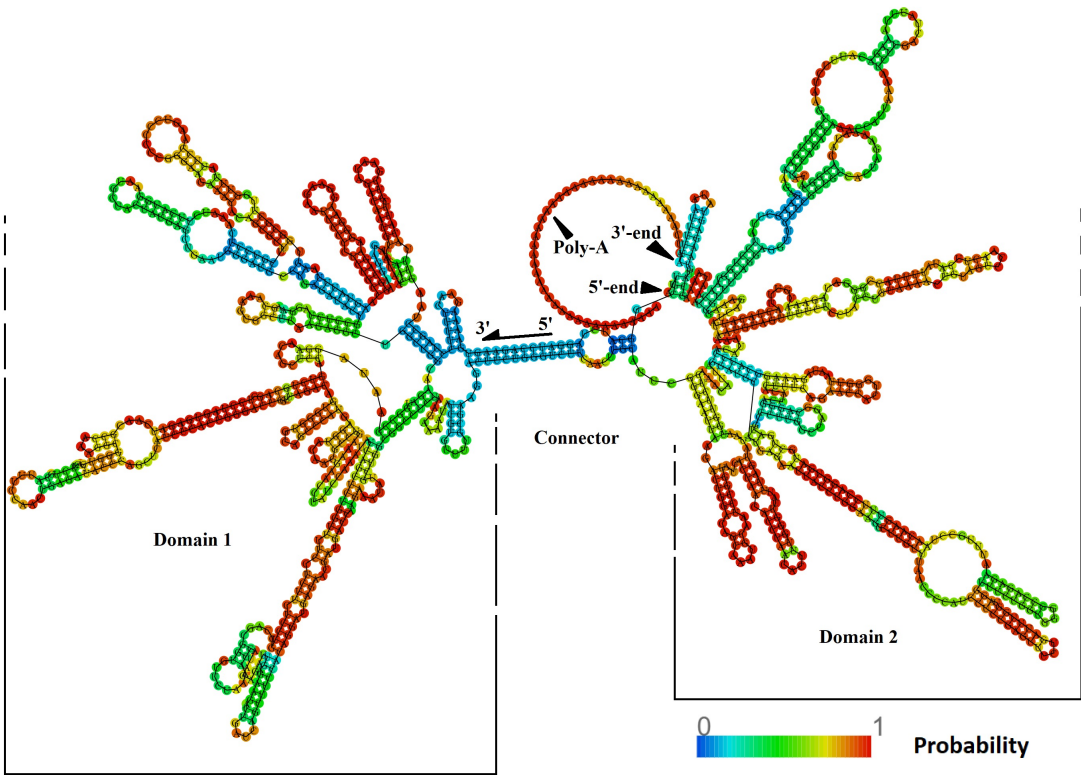
Supplementary Figure 2 : Characterisation of the *I2ALR* poly-adenylation region from the last exon to regions near putative poly-A signals by PCR amplification. A) Using the '*I2ALR* 3'-end primer set 1' a ~500 bp product from poly-A transcripts was generated. **(B)** Combining primer set 1 – set 2 reverse primer, between a classical and non-classical poly-A signal, generated the expected ~560 bp product from poly-A transcripts. Gel ladder with 300, 500 and 600 bp bands marked.).

Supplementary Results Figure 3

A

Energy = -19.26 kcal/mol (9 ideal base pairings) 32 89 5'-AGAGAAGCCUCUGGACAGCUUCU-AGAGUGUGCAGGUUCUGUAUCCUCGCGCAAGGGUAUCCUCUG-3' ITGA2 3'-GAUAAUUGGGUACAC-----AAGACUUUCACA---CCA-GGGAC---GGA-CCGGUCCCCGAGUCGUA-5' LncRNA I2ALR 853 808	Energy = -21.37 kcal/mol (6 ideal base pairings) 3837 3885 5'-AAUGAAGAAUUGUGGGGGUGGGGGAGGUGCGGGGG--CAGGUAGGGAA-3' ITGA2 3'-UUUAGUCUUUGAG-CCUCCUACCCCAUUU-GUUCGGAAGGUCCAUUAGA-5' LncRNA I2ALR 748 710
Energy = -14.81 kcal/mol (9 ideal base pairings) 4548 4589 5'-UAGGAUUUGUUUGGCUACUGGACG-UAACCUAGUGAAUUUCUGAAAGAUAG-3' ITGA2 3'-UGUAAAAAUACACCUACGGAAGAUAAUUGGGUACAC--AAGACUUUCACACCC-5' LncRNA I2ALR 875 835	Energy = -14.01 kcal/mol (7 ideal base pairings) 5506 5518 5'-GACUACCCUGGCGGCGCCGUGG-3' ITGA2 3'-CACCAGGGAC-GGACCGGUUCCC-5' LncRNA I2ALR 828 817
Energy = -15.24 kcal/mol (8 ideal base pairings) 7305 7319 5'-UACAUUCAGGCAUACCCAUUUUAAU-3' ITGA2 3'-CAUAGAGUCCGUA-GGGUAAAGUCGA-5' LncRNA I2ALR 230 217	

B



Supplementary Figure 3. Bioinformatic prediction of I2ALR lncRNA interactions with ITGA2 mRNA and I2ALR secondary structure. (A) “IntaRNA” predicted complementary interactions between *ITGA2* mRNA (7869 bp) and 1112 bp *I2ALR* sequence (combined exons of ENST00000505701.5 + ENST00000503559.1 and uncharacterized 3’ region or U3R). Six potential interactions were predicted with complementary areas of > 6 perfect base pairings, with associated energy scores indicated, ‘|’ and ‘:’ represent Watson-Crick (ideal) and wobble base pairings respectively. (B) “RNAfold” predicted *I2ALR* secondary structure (minimum free energy), predicted structure from the 5 known exons and extended region down to the poly-A signal, base pair probabilities range from 0–1 (dark blue–red), when unpaired the colour denotes probability of being unpaired.