

Supplementary Information

Upcycling Compact Discs for Stretchable and Transient Wearable Electronics

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Alternative Fabrication

To decrease the oval thickness of the metal layer, the processing of the CD can be altered by the removal of the protective layer. The protective layer can be removed by soaking in hydrochloric acid (HCl) or nitric acid (HNO₃). After 10 minutes in non-diluted HCl, or 3.5 min in 3:1 HNO₃:DI water, the protective layer of the CD is removed. The HCl soak produced a thickness of $\sim 70.04 \pm 3.51$ nm and the HNO₃ soak, a thickness of $\sim 18.96 \pm 5.28$ nm (Figure 7E-F). Scanning electron microscope (SEM) images of the removed protective layer reveal the buckles in the metal layer that were engineered into the CDs for data storage, called “lands” and “pits”. The FTIR spectrum of the protective layer after being soaked in HCl and HNO₃ confirms the full removal of the layer, where the characteristic peaks are no longer present, and a metal-carbonyl complex can be seen (Supplementary Fig. 5B). The energy-dispersive X-ray spectroscopy (EDS) of the metal layer after the acetone, HCl, and HNO₃ soak was analyzed (Supplementary Fig. 5B and 9). After the acetone and HCl soak, Ag and Au can be seen within the spectrum at similar concentrations 70.95 and 29.05 wt.% (Supplementary Fig. 9A-D). However, since Ag is dissolved in HNO₃, only Au can be seen by EDS analysis (Supplementary Fig. 9E-F). The ultra-thin metal layer creates difficulties during processing while trying to maintain the structural integrity of the film.

Calibration of Stretchable Resistive Temperature Detector Sensor

Body temperature is a vital sign commonly used in clinical settings to access the current state of a patient’s health. Here, we created upcycled CD electronic (UCDE) temperature sensors that can be used to measure on-skin, real-time temperature. The stretchable resistive temperature detector (RTD) sensor was calibrated by placing it on a hotplate next to a thermocouple and recording the associated 4 probe resistivity with the temperature reading from the thermocouple. The

relationship between the resistivity changes linearly proportional to the temperature that can be denoted by the following equation:

$$R = R_0(1 + \alpha(T - T_0)) \quad (1)$$

where R is the changed resistances, R_0 is the initial resistance, α is the temperature coefficient of resistance (TCR), T is the measured temperature, and T_0 is the initial temperature¹. The results are shown in Figure 3D. The TCR of the UCDE RTD was measured to be 0.0009 °C⁻¹ at 20 °C whereas the TCR of gold is 0.0037 °C⁻¹ at 20 °C. We believe there is a large variation from the TCR of gold because the CD metal isn't a pure gold metal film and due to the ultrathin metal film, surface scattering may occur.

The performance of the calibrated UCDE RTD sensor was compared to the FLIR camera in real-time. Four probe resistance of the UCDE RTD was continuously acquired by a digital multimeter (Keysight, 34460A). Results are presented in Figure 3E. The maximum error of temperature measurement using the UCDE RTD is ± 6.75 °C seen at the hyperpolarization of the temperature curve that could be due to the UCDE recording the temperature more accurately as the heat dissipates through the ambient air.

Electrochemical Performance Evaluation

Chronoamperometry was used to evaluate the sensitivity, dynamic range, and response time of the UCDE amperometric sensors. The electrochemical analysis was conducted with the UCDE reference, counter, and working electrode at room temperature in 1x PBS (pH 7.4). The amperometric response for the enzymatic sensors was monitored by injecting set concentrations of L-lactate or glucose. The oxygen sensor was monitored and modulated by changing the gas concentration in solution by changing the ratio of oxygen and nitrogen gas with a proportioner and

monitoring the dissolved oxygen with a DO meter (Hanna Instruments). The applied potentials were 0.0 V vs. fabricated Ag/AgCl for lactate and glucose sensing and -0.4 V vs. fabricated Ag/AgCl for oxygen sensing.

Open circuit potential (OCP) was used to determine the performance of the pH sensor. The electrochemical cell was the UCDE reference, counter, and working electrode. The (OCP) response was monitored by injecting pH buffer solutions in an aqueous bath while monitoring the pH changes with a pH meter (Hanna Instruments).

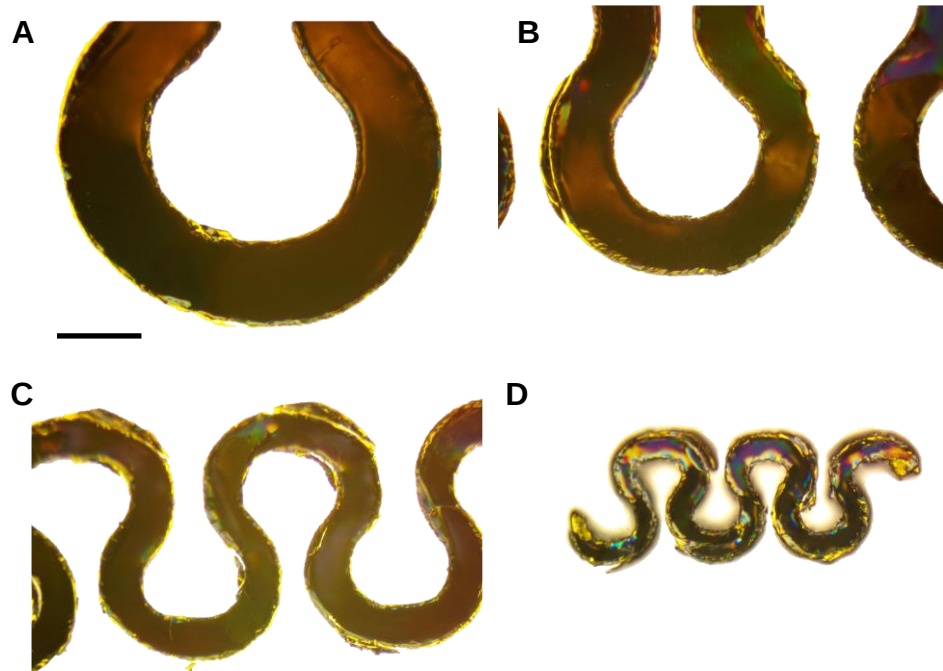
Fabrication of Biodegradable, Transient Electronics

The biodegradable electronics were upcycled through a process illustrated in Supplementary Figure 19. This process required the full removal of the plastic layer on the CD (Supplementary Fig. 4). To remove the plastic layer, the CD was soaked in nitric acid and the plastic layer was delaminated from the metal layer of the CD (Supplementary Fig. 19A and 20). The exposed metal layer from the CD ($\sim 18.96 \pm 5.28$ nm thick) was composed of Au as confirmed by SEM EDX (Supplementary Fig. 7F). This nitric acid soak removed the Ag present in the metal layer leaving behind a film of Au. A solution of 5% PVA was casted and coated with a 50 μ m thick, thin-film bar coater and curing at 80 °C for 1 hr. The PCL-based device was fabricated by reflowing PCL above its melting temperature at 57 °C while coating with a 50 μ m thick, thin-film bar coater and allowing for cooling and recrystallization (Supplementary Fig. 19B). Advanced patterning and structures can be developed with this layout, as PCL has been demonstrated to have strong adhesion to gold and utilized with advanced microfabrication techniques². Next, they were detached from the CD leaving behind a metal-PVA and/or metal-PCL device (Supplementary Fig.

19C). The resistor shape was then patterned into the device by etching with a mechanical cutter and removing the excess (Supplementary Fig. 19D-E).

References

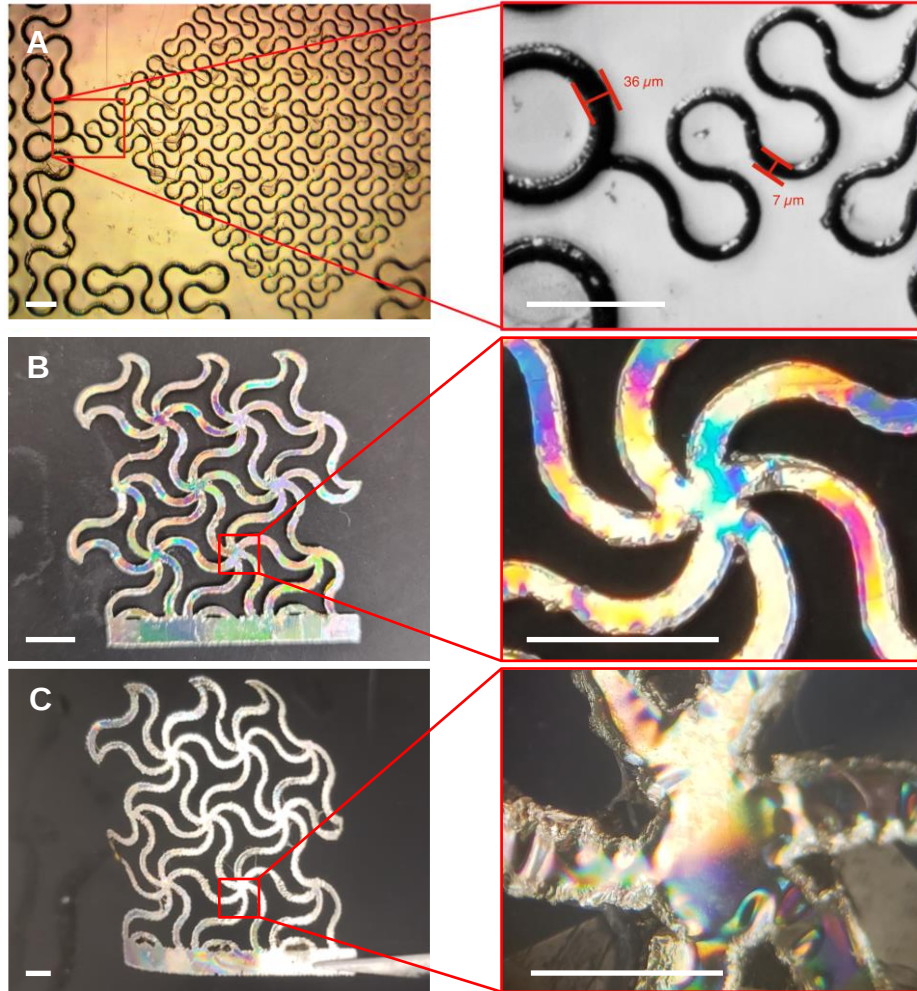
1. Kang S.K., *et al.* Bioresorbable silicon electronic sensors for the brain. *Nature* **530**, 71-76 (2016).
2. Armani D.K., *et al.* Microfabrication technology for polycaprolactone, a biodegradable polymer. *J. Micromech. Microeng.* **10**, 80-84 (2000).



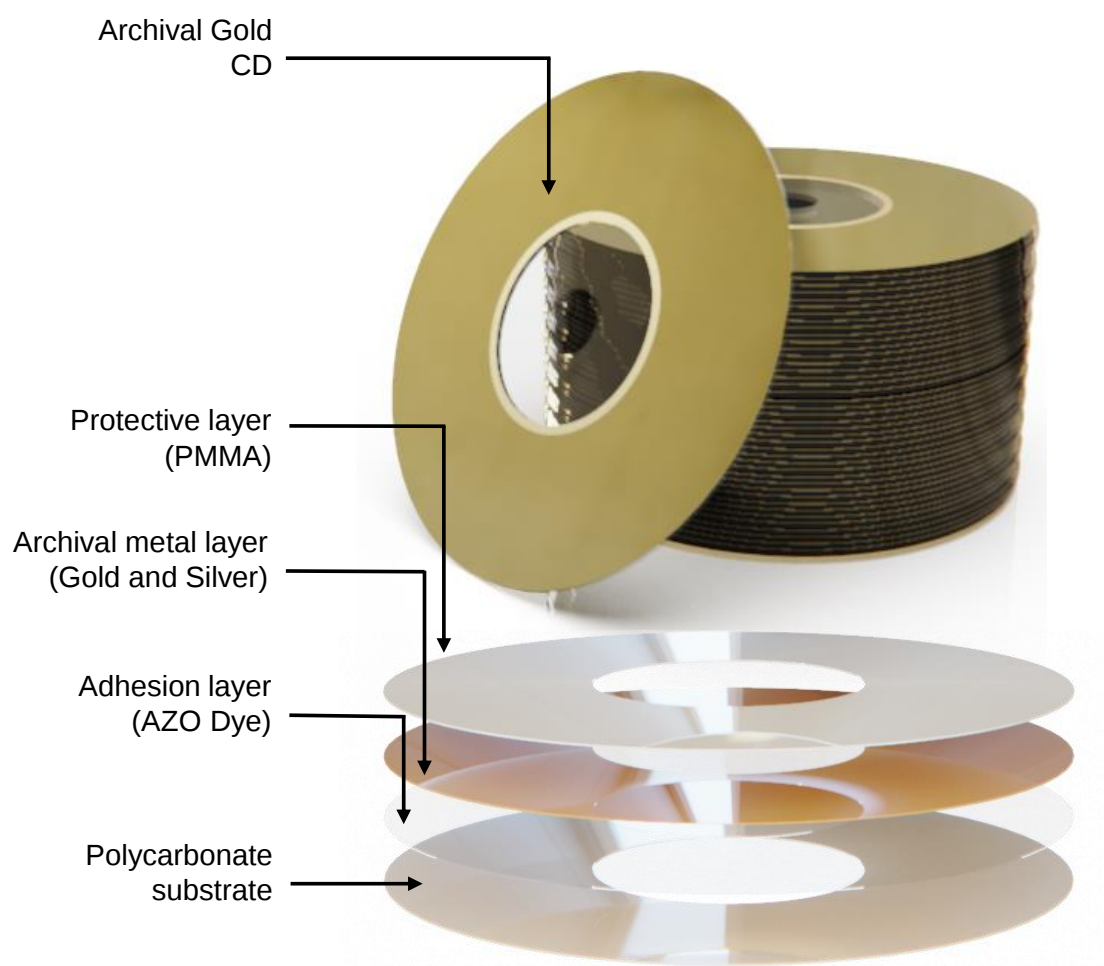
90 **Supplementary Figure 1. Patterning wire resolution.** Various feature sizes: (A) 1 mm (scale
 91 bar, 1 mm), (B) 0.75 mm, (C) 0.5 mm, and (D) 0.25 mm.



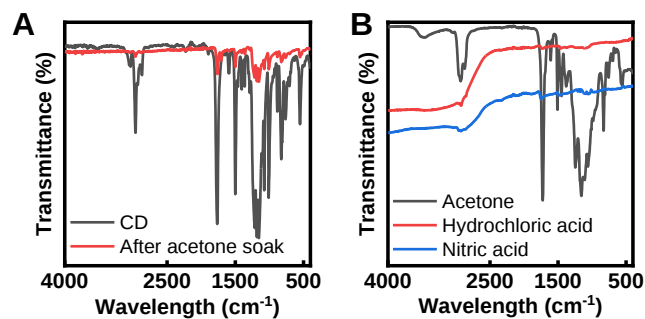
92 **Supplementary Figure 2. Photograph of the UCDEs during stretching (scale bar 1 mm).**



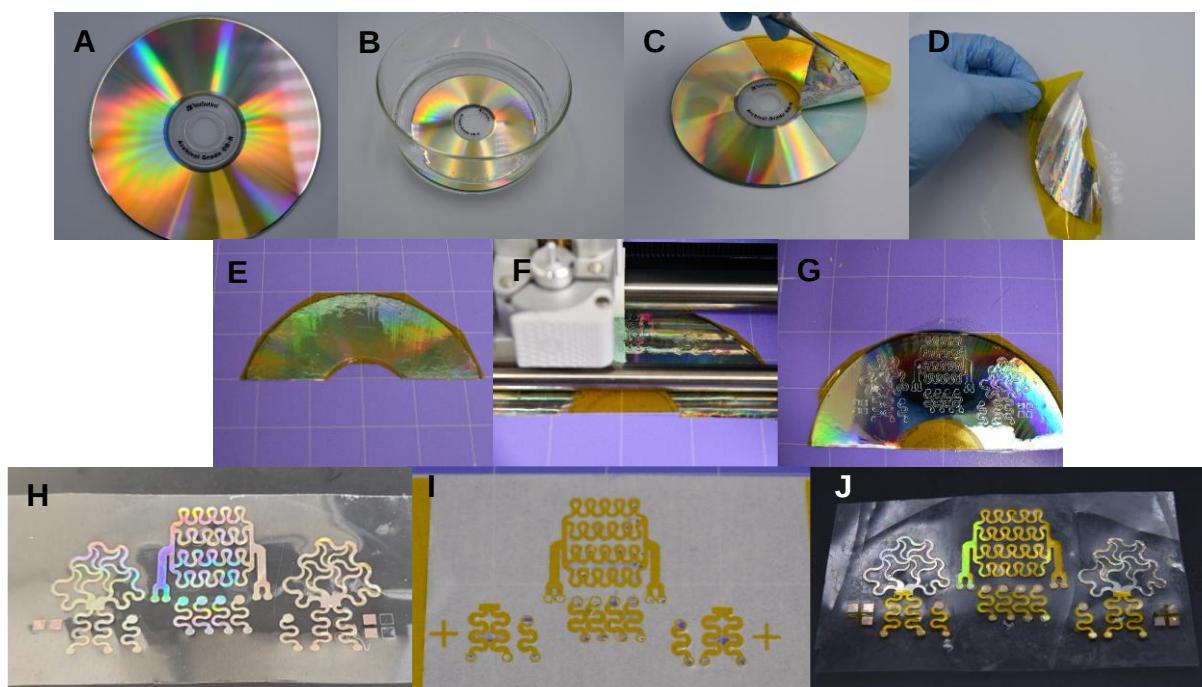
Supplementary Figure 3. Various patterning methods for the UCDEs. (A) Photolithography (scale bar, 200 μm). (B) Mechanical cutter (scale bar, 4 mm) (C) CO₂ laser engraver (scale bar, 2 mm).



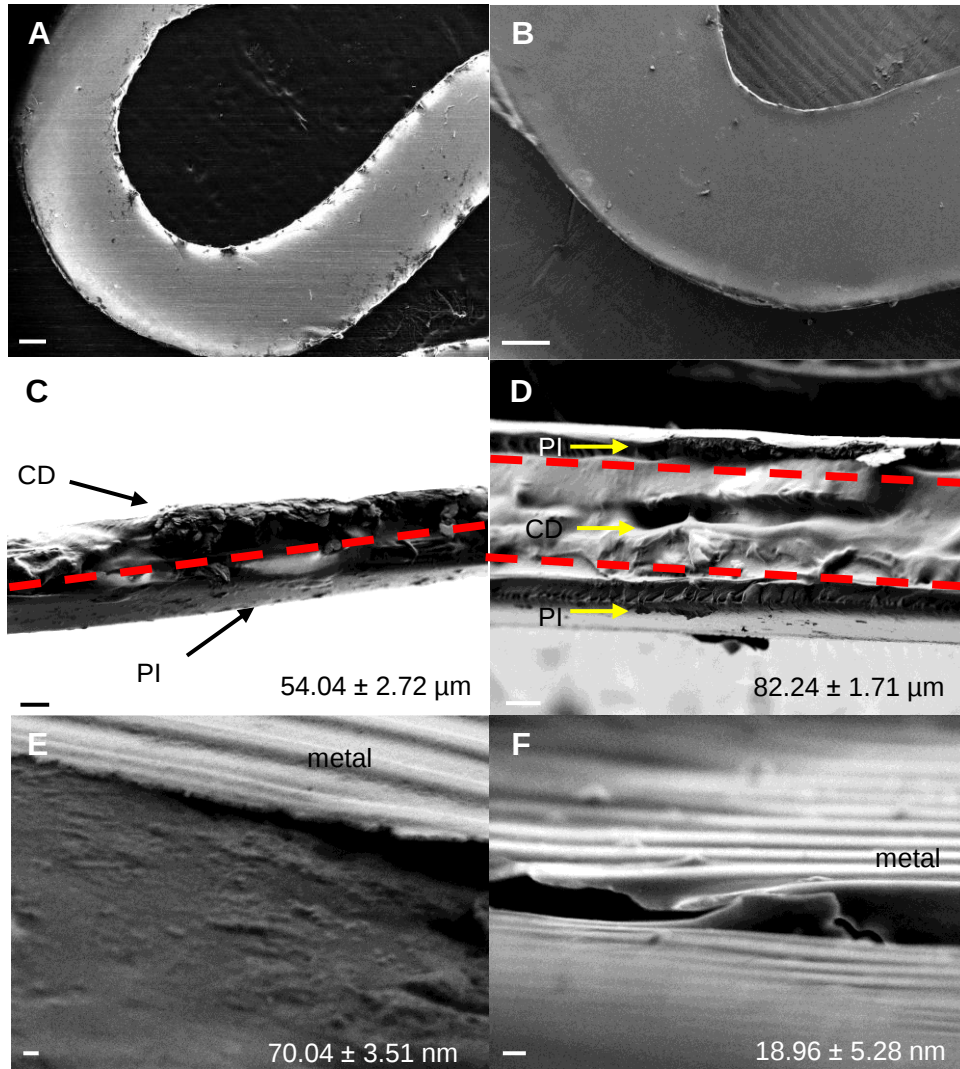
96 **Supplementary Figure 4. Structural schematic of an archival gold CD.**



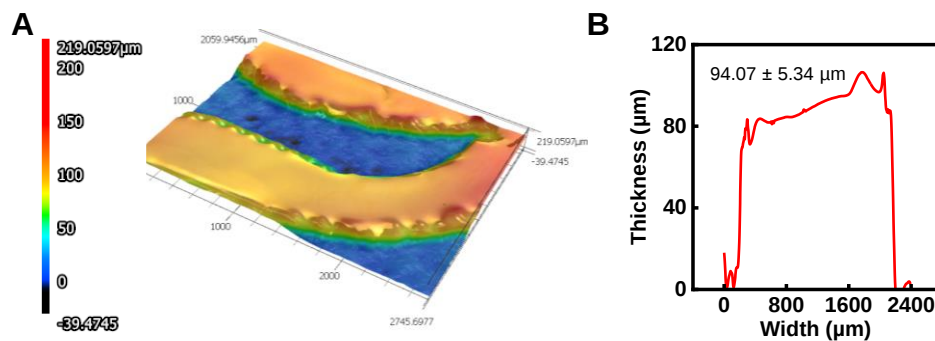
97 **Supplementary Figure 5.** (A) FTIR spectrum from the substrate polycarbonate layer, before and
 98 after soaking the CD in acetone. (B) FTIR spectrum of the protective layer after soaking in acetone,
 99 hydrochloric acid, and nitric acid.



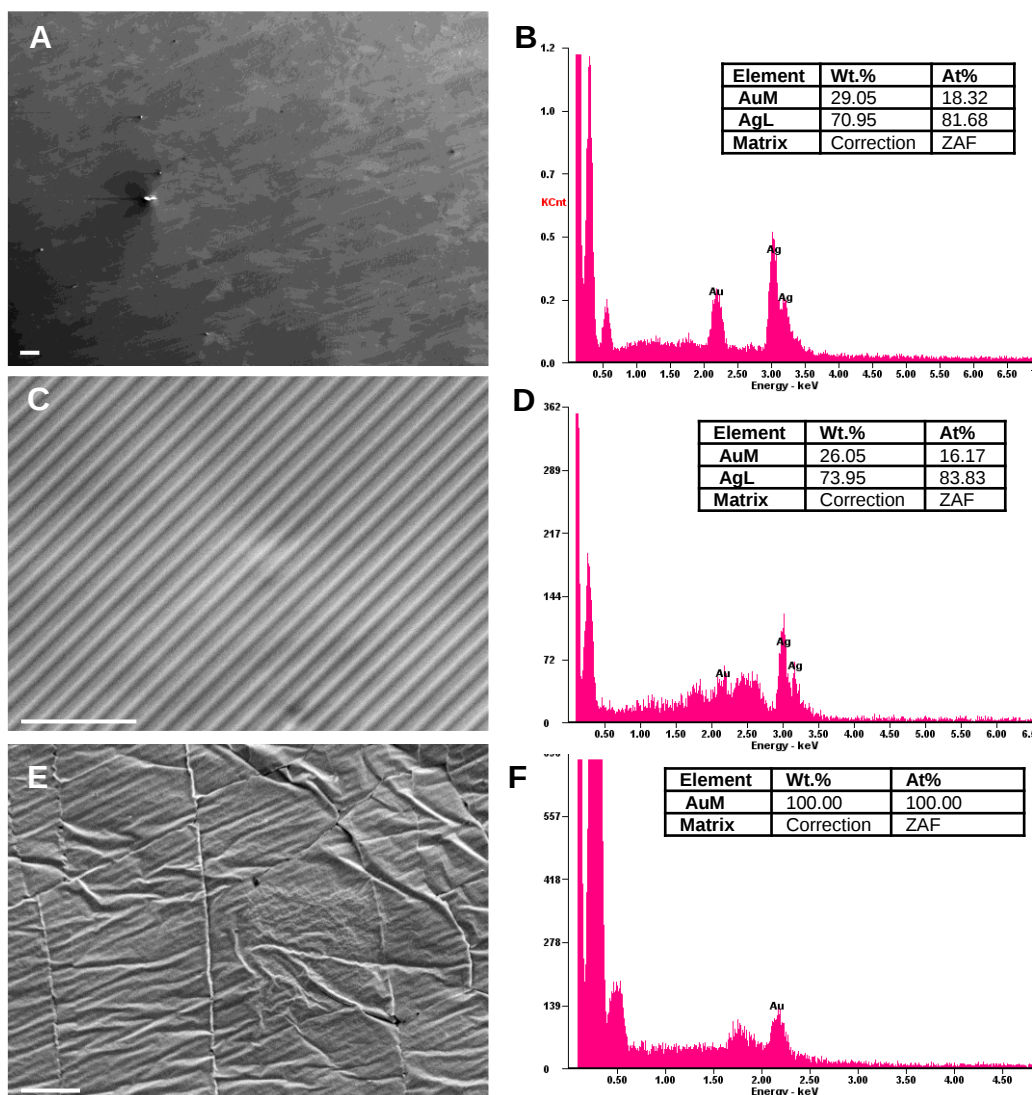
Supplementary Figure 6. The fabrication process of the UCDEs. (A) Archival gold CD. (B) Soaked in acetone. (C) Harvesting the metal layer from the CD with PI tape. (D) Image of the harvested metal layer. (E) The PI side adhered to tattoo paper and then placed on the cutting mat. (F) Patterning with the cutting machine. (G) Post patterning. (H) Removal of excess. (I) Image of the insulation layer post cutting and after removal of the excess. A similar process is administered with the PI tape from steps E–H. The PI tape adhered onto water-soluble tape as a temporary substrate. (J) Full UCDE device. The PI tape is laminated over the patterned metal layer.



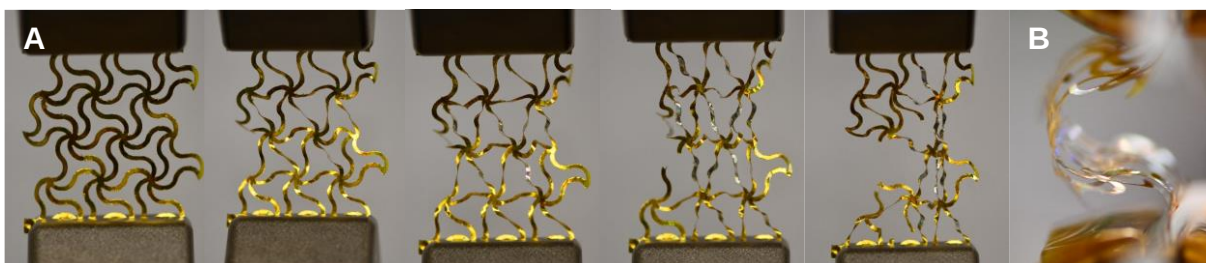
Supplementary Figure 7. SEM images of UCDEs. (A) The metal layer, post patterning (scale bar, 200 μm). (B) Full encapsulation with PI (scale bar, 200 μm). (C) Cross-sectional image of the PI and CD metal (scale bar, 20 μm). (D) Cross-sectional image after full encapsulation with PI (scale bar, 20 μm). Metal layer remaining after (E) hydrochloric acid and (F) nitric acid soak (scale bar, 200 nm). The thickness of the metal layer is denoted in the bottom right of the image.



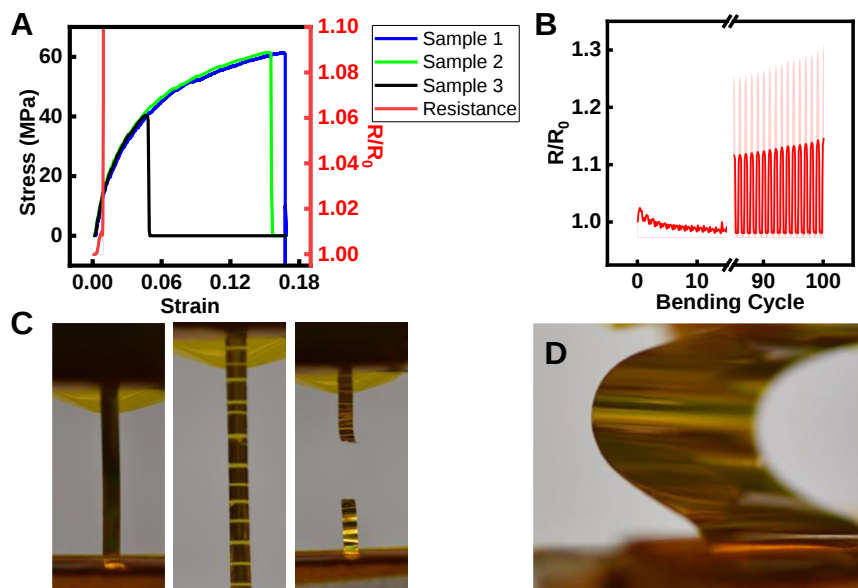
112 **Supplementary Figure 8. Optical profilometer analysis of UCDEs post full insulation. (A)**
 113 **Three-dimensional profile of UCDEs. (B) Thickness profile vs. width of UCDEs.**



114 **Supplementary Figure 9. SEM images and EDS analysis of harvested metal films from CD.**
 115 Acetone soak (A) SEM image (scale bar, 100 μm) and (B) EDS spectrum. Hydrochloric acid soak
 116 (C) SEM image (scale bar, 10 μm) and (D) EDS spectrum. Nitric acid soak (E) SEM image (scale
 117 bar, 10 μm) and (F) EDS spectrum.



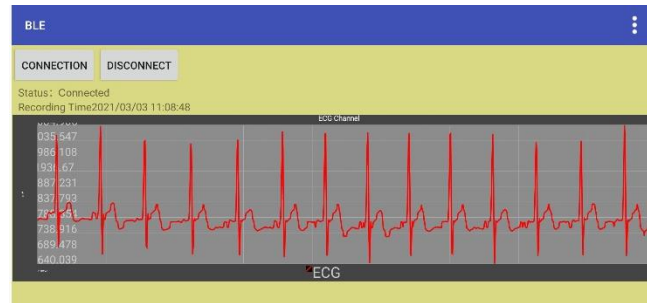
118 **Supplementary Figure 10. Mechanical response of patterned CD, UCDEs.** (A) Images of
 119 tensile testing. (B) Image of bending.



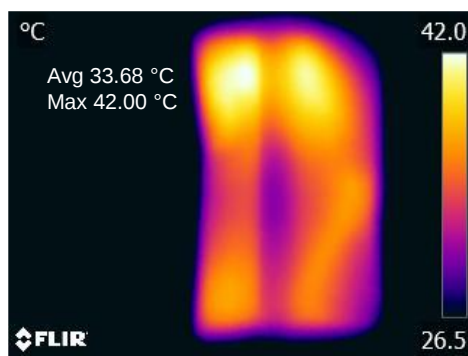
120 **Supplementary Figure 11. Mechanical properties of unpatterned CD electronics.** (A) Stress
 121 and strain vs. electrical performance and (B) resistance vs. cyclic bending of unpatterned samples.
 122 Images of the unpatterned samples under (C) stretching and (D) bending.

123 **Supplementary Table 1. Mechanical properties pattern and unpatterned samples.**

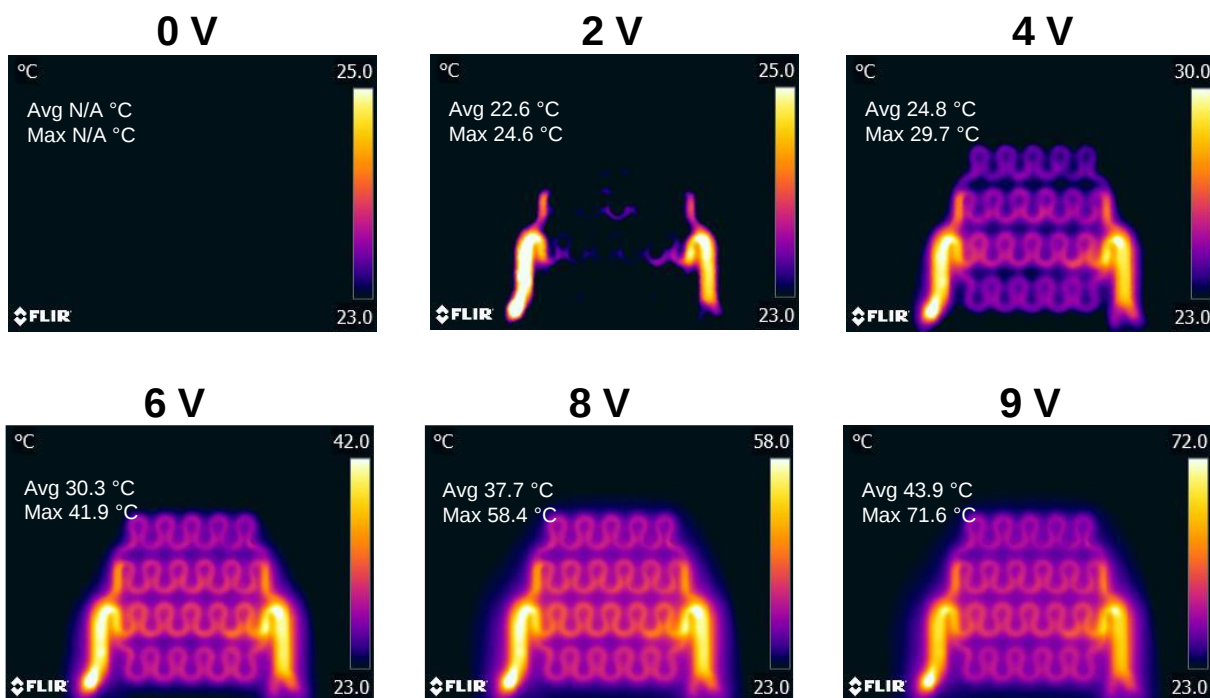
Sample	Young's Modulus	Yield Strength	Ultimate Tensile Strength	Yield Strength Elongation (%)	Elongation (%)
Patterned UCDEs	5.59 ± 0.16 MPa	0.95 ± 0.07 MPa	0.99 ± 0.06 MPa	62.36 ± 1.81	129.24 ± 3.43
Unpatterned CD Electronics	1.88 ± 0.02 GPa	12.85 ± 0.91 MPa	54.47 ± 6.99 MPa	0.09 ± 0.05	12.39 ± 3.76
PI Tape	3.60 ± 0.60 GPa	32.45 ± 8.80 MPa	0.16 ± 0.02 GPa	0.88 ± 0.23	24.40 ± 6.52



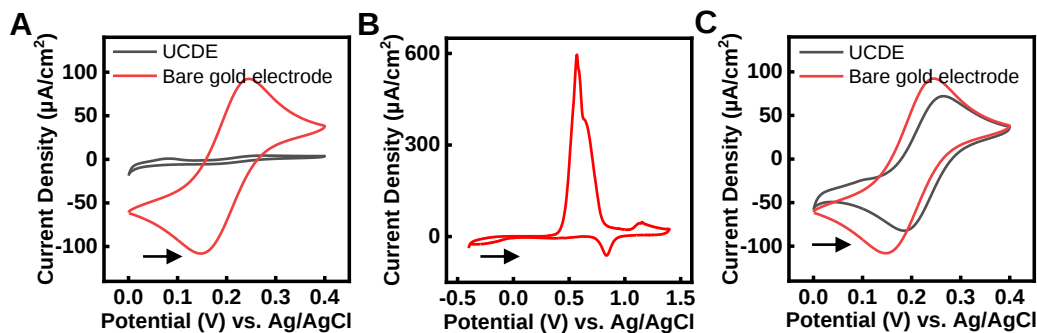
124 **Supplementary Figure 12. Screenshot of the smartphone application, recorded ECG signal.**



125 **Supplementary Figure 13. IR image of commercially available hand warmers (Hot Hands).**



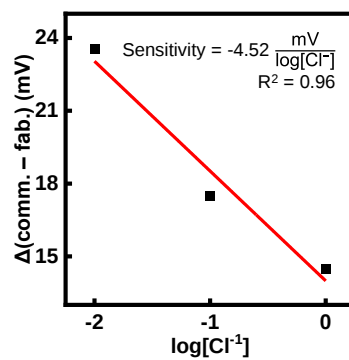
126 **Supplementary Figure 14. Larger, stretchable UCDE heater (3.5 cm width).**



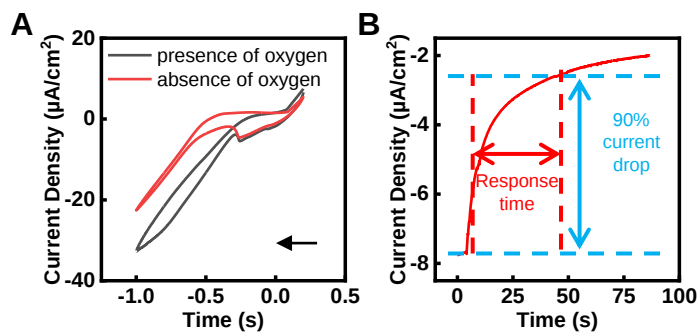
Supplementary Figure 15. Characterization of the modified UCDE electrode before and after electrochemical cleaning. (A) Cyclic voltammetry at 50 mV/s of UCDEs (before cleaning) and bare gold electrode vs. Ag/AgCl (1 M KCl) in PBS (pH 7.4) with 5 mM $K_3Fe(CN)_6$. (B) Electrochemical cleaning of UCDE vs. Ag/AgCl (1 M KCl) in 0.1 M H_2SO_4 at 25 mV/s. Cyclic voltammetry at 50 mV/s of UCDEs (after cleaning) and bare gold electrode vs. Ag/AgCl (1 M KCl) in PBS (pH 7.4) with 5 mM $K_3Fe(CN)_6$.

133 **Supplementary Table 2. The characteristics of the fabricated UCDE Ag/AgCl reference**
134 **electrode vs. a commercial Ag/AgCl (1 M KCl) reference electrode.**

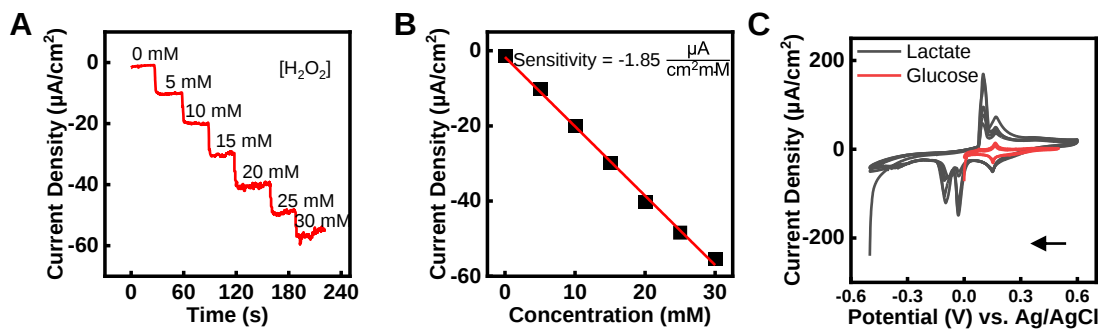
1 mM K ₃ Fe(CN) ₆				
1 M KCl				
	E _{1/2} (mV)	Δ(comm – fab) (mV)	ΔE (mV)	I _p a/I _p c
Commercial	261.00		74.00	1.54
Fabricated	246.50	14.50	83.00	1.17
1 mM K ₃ Fe(CN) ₆				
0.1 M KCl				
	E _(1/2) (mV)	Δ(comm – fab) (mV)	ΔE (mV)	I _p a/I _p c
Commercial	230.50		87.00	1.98
Fabricated	213.00	17.50	90.00	0.98
1 mM K ₃ Fe(CN) ₆				
0.01 M KCl				
	E _(1/2) (mV)	Δ(comm – fab) (mV)	ΔE (mV)	I _p a/I _p c
Commercial	217.00		106.00	1.13
Fabricated	193.45	23.55	101.10	0.70



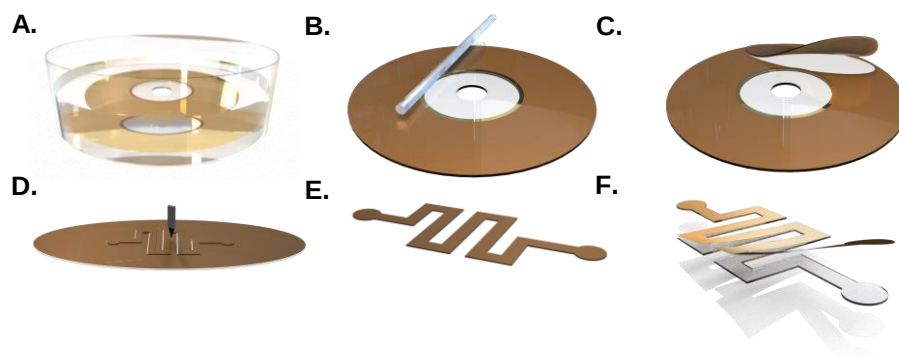
135 **Supplementary Figure 16. Calibration curve of fabricated vs. commercial reference**
 136 **electrode.**



137 **Supplementary Figure 17. Characterization of the UCDE oxygen electrode.** (A) Cyclic
 138 voltammetry of the UCDE oxygen electrode in the presence and absence of dissolved oxygen. (B)
 139 Sensor response time from oxygen saturated state to depleted state.



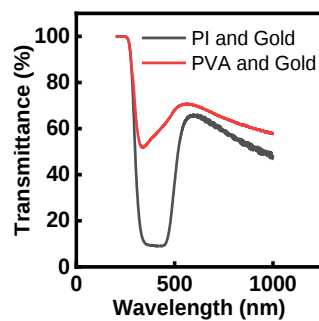
Supplementary Figure 18. Characterization of the Prussian Blue modified UCDE electrode.
 (A) The amperometric $i-t$ curves of the Prussian Blue, H_2O_2 detection sensor. (B) Calibration curve of the current density vs. H_2O_2 concentration. (C) Electrochemical deposition of Prussian Blue via cyclic voltammetry vs. Ag/AgCl (1M KCl).



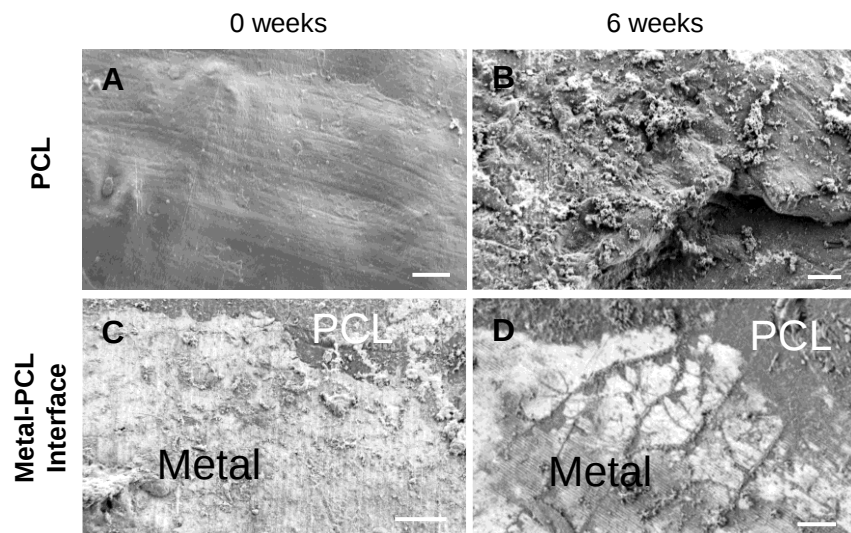
Supplementary Figure 19. Upcycling CDs into Transient, Biodegradable Electronics. (A) Soaked in nitric acid. (B). Bar coated with a biodegradable polymer and cured. (C) Peeled off biodegradable polymer and gold layer from the CD. (D) Patterned with a mechanical cutter. (E) Removed excess and yielded a biodegradable device. (F) The two-layer layout of the biodegradable device.



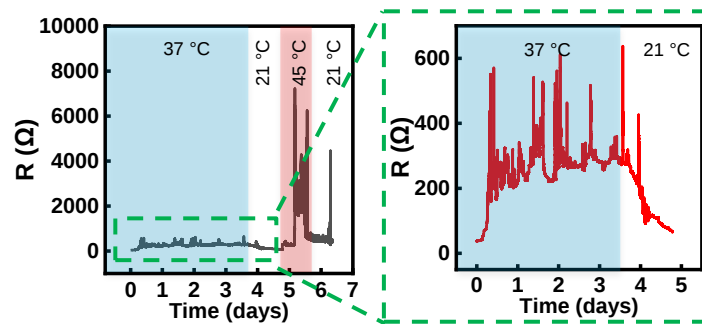
149 **Supplementary Figure 20. CD after the nitric acid soak.**



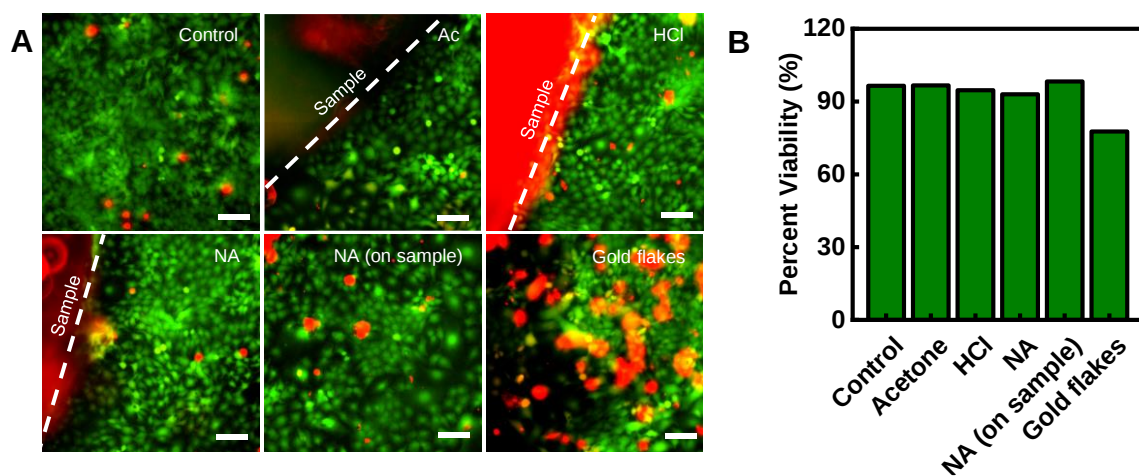
150 **Supplementary Figure 21. UV-Vis of the biodegradable PVA-based UCDE device.**



Supplementary Figure 22. SEM images of PCL resistor degradation in PBS (7.4 pH) at 37 °C. PCL substrate at (A) 0 weeks and (B) 6 weeks. Metal-PCL interface at (C) 0 weeks and (D) 6 weeks. Scale bar 10 μm .



154 **Supplementary Figure 23. Long-term resistance stability of transient UCDE on PCL matrix.**
 155 Soaked in PBS (pH 7.4) at 37 °C and 45 °C.



Supplementary Figure 24. Biocompatibility of UCDEs. (A) Fluorescent imaging of live/dead stained HaCaT cell cultured for 7 days (scale bar, 100 μ m). Sample group of the soaking method for UCDEs: Acetone (Ac), Hydrochloric acid (HCl), Nitric acid (NA), Na (image directly on the sample), and gold flakes. (B) Cell viability at day 7. Analysis produced by fluorescent imaging viability counting.